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The basolateral nucleus of the amygdala executes the parallel processes of avoidance and palatability in the retrieval of conditioned taste aversion in male rats

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2 The basolateral nucleus of the amygdala executes the parallel processes of avoidance and palatability in
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4

5 Abbreviated title:

6 BLA avoidance and palatability on CTA retrieval

7

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49

50 **Abstract**

51 Conditioned taste aversion (CTA) is an essential behavior for animal survival. Conditioned animals show
52 avoidance and decreased palatability to a conditioned stimulus (CS) on CTA retrieval. In this study, we
53 aimed to determine whether the basolateral nucleus of the amygdala (BLA) is involved in CTA retrieval
54 and whether avoidance and palatability in CTA retrieval are processed in the BLA. We developed an
55 experimental chamber for time-course analysis of the behavior to approach a spout and lick a CS. In this
56 experimental chamber, we analyzed the behavior of male rats following microinjections of
57 gamma-aminobutyric acid (GABA_A) receptor agonist muscimol or saline into the BLA. The rats showed
58 two types of approach behavior: they either (1) approached and licked the spout or (2) approached but did
59 not lick the spout. Muscimol injection into the BLA decreased the frequency of the latter type of approach
60 behavior, indicating that BLA inactivation reduced avoidance to the CS. The muscimol injection into the
61 BLA also significantly increased the consumption of the CS. Lick microstructure analysis demonstrated
62 that intra-BLA muscimol significantly increased licking burst number and size, indicating that BLA

63 inactivation attenuated aversion to the CS as large burst licking is an indicator of high palatability. These
64 results suggest that the increase in CS consumption with intra-BLA muscimol injection was due to
65 alterations in approach and aversive responses to the CS. Therefore, we conclude that the BLA plays an
66 essential role in CTA retrieval by parallel processing of avoidance and palatability.

67

68 **Significant statements**

69 To understand the role of the basolateral nucleus of the amygdala (BLA) in the retrieval of conditioned
70 taste aversion (CTA), we developed a new behavioral paradigm that overcomes the limitations of
71 traditional behavioral procedures in CTA studies. In male rats, we measured time stamps of the approach
72 behavior to a conditioned stimulus (CS) and licking of a spout. BLA inactivation not only increased CS
73 consumption but also decreased avoidance and aversion to the CS. These results reveal that the BLA is
74 essential for CTA retrieval by processing two distinct behavioral responses in parallel: avoidance and
75 palatability. We further demonstrate that multi-dimensional analysis of animal behavior is an important
76 factor for elucidating brain function.

77

78 **Introduction**

79 Food can be contaminated with harmful substances. Malaise after eating contaminated food induces
80 conditioned taste aversion (CTA) through the association between food and taste as a conditioned
81 stimulus (CS) and malaise as an unconditioned stimulus (US). Conditioned animals show suppression of
82 CS consumption, which is induced by avoidance (Parker, 2003), and decrease in palatability (Spector et
83 al., 1988) to the CS on CTA retrieval.

84 The suppression of CS consumption on CTA retrieval is attenuated by microinjection of the
85 benzodiazepine agonist midazolam or the AMPA receptor antagonist
86 2,3-dihydroxy-6-nitro-7-sulfamoyl-benzo[f]quinoxaline-2,3-dione (NBQX) (Yasoshima and Yamamoto,
87 2005; Garcia-Delatorre et al., 2014). CS inhibits electrophysiological activities in 76% of recorded
88 basolateral nucleus of the amygdala (BLA) units (Kim et al., 2010) and induces ERK activation in the
89 BLA (Lin et al., 2010). However, there are controversial reports stating that pharmacological BLA
90 inactivation may not impair CTA retrieval (Bahar et al., 2004; Garcia-Delatorre et al., 2010;
91 Garcia-Delatorre et al., 2014), and an intraoral infusion of an aversive CS does not alter Fos-like
92 immunoreactivities in the BLA (Yamamoto et al., 1997). These contradictory findings suggest that more
93 evidence is required to determine the role of the BLA in CTA retrieval.

94 We previously demonstrated that a CS activates the neuronal projections from the BLA to the nucleus
95 accumbens core (NAc), the anterior part of the bed nucleus of the stria terminalis (BNST), and the
96 central nucleus of the amygdala (CeA) (Inui et al., 2013). The projections from the NAc to the ventral
97 pallidum (VP) mediate decreased palatability of a conditioned aversion to saccharin (Inui et al., 2007;
98 Inui et al., 2009; Inui et al., 2011; Inui and Shimura, 2014, 2017). The projections from the BLA to the

99 BNST and the CeA exert an effect on fear and anxiety (Walker et al., 2009). Although the unique
100 functions of these targets—NAcC, BNST, and CeA—suggest multiple roles for the BLA in behavioral
101 expression, there is no evidence showing that the BLA is involved in either avoidance or palatability to a
102 CS on CTA retrieval.

103 Previous studies on the BLA in CTA retrieval traditionally have employed either intake or taste reactivity
104 tests. The intake test measuring consumed fluid volume provides direct evidence of voluntary fluid
105 consumption and indirect evidence of avoidance and palatability because other factors, such as motivation
106 and post-ingestive effects, can affect consumption. Although a taste reactivity test can evaluate
107 palatability by analyzing animal oromotor responses to intraorally infused fluid (Grill and Norgren, 1978;
108 Spector et al., 1988), investigating the avoidance response is unsuitable because the animals receive
109 passive intra-oral fluid infusion. To overcome the limitations of these traditional behavioral procedures,
110 we developed a new apparatus for multi-dimensional behavior analysis. This apparatus measures the
111 pattern of licking displayed by animals during fluid consumption. The ingestive behavior for animals
112 consuming fluids consists of sustained runs of rapidly occurring rhythmic licks—referred to as a
113 burst—separated by pauses of varying lengths. Animals display different burst size (i.e. number of licks in
114 a burst) between palatable and aversive taste solutions (Davis and Smith, 1992; Davis and Perez, 1993;
115 Hsiao and Fan, 1993; Davis, 1996; Spector and St John, 1998; Monk et al., 2014). We applied these
116 microstructural analyses of licking to evaluate palatability in animals drinking a CS voluntarily. The
117 experimental chamber was also used to assess the avoidance produced by the establishment of CTA by
118 measuring the approach behavior of how the animals attempted to come close to the CS by detecting the
119 tip of the animal snout (Parker, 2003).

120 Using the experimental chamber and microinfusions of GABA_A receptor agonist muscimol into the BLA,
121 we aimed to determine whether the BLA is involved in CTA retrieval and, if so, whether avoidance or
122 palatability on CTA retrieval is processed by BLA neurons.
123

124 **Materials & methods**

125 *Subjects*

126 Twenty-three male Wistar rats (CLEA Japan, Inc., Osaka, Japan) weighing 250-280 g on arrival were
127 individually housed in a temperature- and humidity-controlled colony room with a 12-h light/dark cycle
128 (7:00 A.M. lights on). Water and food (MF, Oriental Yeast Co., Ltd., Tokyo, Japan) were available ad
129 libitum in the home cage, unless otherwise specified. All animals were handled in accordance with the
130 procedures outlined in the “Guide for the Care and Use of Laboratory Animals” (National Institutes of
131 Health Guide), and all experiments were approved by the Institutional Committee on Animal Research of
132 Graduate School of Human Sciences, Osaka University.

133

134 *Surgery*

135 Following acclimation in the colony room for at least one week, guide cannulae were bilaterally
136 implanted into the BLA of the rats. Under isoflurane anesthesia (induction 5%; maintenance 1.5-2%),
137 their heads were fixed on a stereotaxic apparatus (SR-6, Narishige Scientific Instrument Laboratory,
138 Tokyo, Japan). The scalp was incised, and the skull was exposed. Small holes were drilled above the
139 targeted brain region. The coordinates were as follows: anteroposterior = -2.8 mm (posterior to bregma),
140 mediolateral = ± 5.0 (lateral to midline), dorsoventral = -7.5 mm (ventral to bregma). The guide cannulae
141 (C315G, Plastics One Inc., Roanoke, VA, USA) were lowered into the holes and secured into the skull
142 using metal anchor screws and dental resin. After suturing of the wound, an analgesic (carprofen, 4.4
143 mg/kg, s.c.) and antibiotic (cefazolin, 0.5 mg/kg, i.m.) were administered. The rats recovered for at least
144 one week with ad libitum water and food before the behavioral experiments.

145

146 *Apparatus*

147 All behavioral experiments were performed in a custom-made chamber (Fig. 1) that consisted of a start
148 box, an arena, and a bay window (Fig. 1a). Animals were initially put in the start box ($100 \times 250 \times 400$
149 mm) and allowed to move to the arena ($40 \times 25 \times 40$ cm) through a guillotine door (100×100 mm)
150 opened by an experimenter (Fig. 1b). The bay window was on the opposite side of the guillotine door. The
151 size of the arena-side bay window was 50 mm high and 40 mm wide, while the outer-side of the box was
152 30 mm high and 40 mm wide, indicating that the bay window was tapered off and fitted to the shape of
153 the rat's head (Fig. 1c). There were three sets of infrared LEDs and infrared sensors that were parts of the
154 passage detection sensor circuit (SY-852, KYOHRITSU ELECTRONIC INDUSTRY Co., Ltd., Osaka,
155 Japan) over and under the bay window (Line 1, 2, and 3 in Fig. 1d). The transparent wall of the bay
156 window allowed the infrared rays from the LEDs to reach the sensors (Fig. 1e). The three sets of LEDs
157 and sensors were located at intervals of 2.5 cm. By analyzing which of the three sets of LED and sensors
158 were shut off, we could determine the depth of entry of the rat's head into the bay window. The shutting
159 off of the infrared light between the LEDs and sensors triggered a square-wave pulse function generator
160 (Akizuki Denshi Tsusho Co., Ltd., Tokyo, Japan). A data logger (TUSB-S03CN3BZ, Turtle Industry Co.,
161 Ltd., Ibaraki, Japan) was used to collect the data of the square-wave pulses with a 10-Hz sampling rate
162 that indicated the timing of entry of the rat's head into the bay window.

163 In front of the bay window, a drinking tube was placed on a plastic holder. The drinking tube consisted of
164 a metal spout and a conical tube (Fig. 1d, e). The metal spout was connected to a touch sensor circuit
165 (SW-104, Kyohritsu Electronic Industry Co., Ltd., Osaka, Japan) via a metal clip and an electric wire. The

166 touch sensor produced square pulse waves, but the current was too small to generate inputs to the data
167 logger. Thus, the currents of the square pulse were amplified using an isolator (M2Y-6A-R2, M-System
168 Co., Ltd., Osaka, Japan) and collected via the data logger for passage detection.

169

170 *Procedures of behavioral experiment*

171 Habituation

172 The animals were acclimated to the chamber for 10 min on the first day in the start box. As the
173 guillotine door was opened during the session, they could move in the chamber freely. The water
174 deprivation schedule was started in the evening of the same day.

175

176 Training

177 Animals deprived of water for 20 hours were initially put in the start box, and after 30 s an
178 experimenter opened the guillotine door to allow the animals to move into the arena. After a rat
179 completely moved to the arena, the guillotine door was closed. The session started at the opening of the
180 guillotine door. The animals were allowed to drink water during a 30 min session. One hour after the end
181 of the session, the rats were given food and water in their home cages until the next water deprivation
182 period. This training was repeated for 4 days.

183

184 Conditioning

185 In this study, we expected the rats to retain aversion through the tests; we planned to examine the
186 establishment of CTA on the first test and the effects of drug injections in second. We previously paired a

187 CS with a US and found that some rats showed a rapid extinction of CTA after the first presentation of the
188 CS (Inui et al., 2007). We needed rats to retain aversion through at least two test trials because we planned
189 to validate the establishment of CTA on the first test and examine the drug effects on the second test. We
190 also expected the conditioning procedure to not change the baseline of animal consumption behavior. In
191 our preliminary study, we found that double conditioning after familiarizing with a CS is an ideal
192 experimental procedure. Therefore, in this study, all the animals were pre-exposed to a 5 mM saccharin
193 solution as a CS for three consecutive days (Days 6-8), followed by two pairings of the CS with an i.p.
194 injection of 0.15 M lithium chloride (LiCl) (2% B.W.) as a US on Days 9 and 11. Five minutes after the
195 end of the session, the US was administered in the home cage, and two hours later, the animals were
196 given ad lib food and water overnight for recovery. On Days 10 and 12, water deprivation was re-started
197 20 hours before the start of the next day sessions.

198

199 Test

200 The water sessions on Days 13 and 14 assessed that the conditioning procedures did not cause changes in
201 the baseline of animal consumption behavior. We confirmed the establishment of CTA on the first test
202 (Test 1, Day 15). All the rats were microinjected with the vehicle (saline) into the bilateral BLA, 5 min
203 before the start of the sessions. They were presented with the CS in the chamber. After Test 1, the rats
204 were divided into two groups that showed no significant differences in the CS intake (total number of
205 licks) in the first conditioning (Conditioning) test, Test 1. In the second test (Test 2, Day 17), one of the
206 groups received bilateral BLA microinjections of GABA_A receptor agonist muscimol, which inhibits

207 neural activation. A control group was microinjected with saline. The experimental procedures of Test 2
208 were identical to those used in Test 1, except for the injected drugs.

209

210 *Microinjection of drug*

211 Muscimol (M1523, Sigma-Aldrich Japan, Tokyo, Japan) was dissolved in sterilized saline (Otsuka
212 Normal Saline: Otsuka Pharmaceutical Factory, Inc., Tokushima, Japan) as a vehicle at a dose of 100
213 ng/0.5 μ l. We used injector cannulae (C315I, Plastics One Inc., Roanoke, VA, USA), which extended 1
214 mm from the tip of the implanted cannulae guide. The injector cannulae were connected to 10- μ l of
215 Hamilton gastight syringes (1701N; Hamilton Company, Reno, NV, USA) via polyethylene tubes (PE10;
216 Becton, Dickinson and Company, Sparks, MD, USA) and Viton tubes (JB-30; Eicom Corporation, Kyoto,
217 Japan) filled with Fluorinert® (FC-40; Sumitomo 3 M, Tokyo, Japan). The syringes were mounted on a
218 syringe pump (KDS210; KD Scientific, Holliston, MA, USA). Muscimol or saline was injected at a rate
219 of 0.25 μ l/min for 2 min (total volume was 0.5 μ l/side). The dose of muscimol was selected from previous
220 studies, which showed inhibition of electrophysiological activity within a 1 mm radius of the injection
221 area (Arikan et al., 2002) and impairment of behavioral performance (Wang et al., 2006; Piette et al.,
222 2012).

223

224 *Behavioral analysis*

225 The outputs from the sensors were plotted in raster images (Fig. 2). These plots show both the entry of the
226 rat's head into the bay window and the licking pattern on an identical time scale. Based on these data, we
227 categorized the approach and licking behaviors as follows.

228

229 Entry

230 We defined Lines 1, 2, and 3 as the combination of the infrared LEDs and sensors at the bay window (Fig.
231 1d), with Line 1 on the side of the arena. We also defined “Entry” as the shutting-off, meaning that the tip
232 of rat’s snout entered the bay window. The rats showed two types of entries (Fig. 3a). The entrance of
233 their heads into the bay window with subsequent licking to the spout is referred to as the “Entry-Lick.”
234 Alternatively, “Entry-Stop” was the event on which the rats entered their heads into the bay window but
235 did not lick at the spout (their head stopped and stayed at the bay window).

236

237 Licking

238 There are several lines of evidence, which suggest that microstructural analysis of animal’s licks to taste
239 solutions reveal their palatability. Davis and Perez (1993) have defined “burst” as three or more licks with
240 <250 ms inter-lick interval (ILI) between them. They showed that higher palatability of taste solutions
241 produces increased number of bursts, burst duration (time from the first lick to last one in each burst), and
242 burst size (number of licks in each burst). They have also defined inter-burst interval (IBI) as ≥ 250 ms
243 and <500 ms ILI, and inter-cluster interval (ICI) as ≥ 500 ms or <1000 ms ILI, and Pause as ≥ 1000 ms ILI.
244 In this study, as the 10 Hz sampling rate produced a 100-ms time window, we have defined ILI as <200
245 ms, IBI as ≥ 200 and <500 ms, and other parameters were same as defined by Davis and Perez (1993) (Fig.
246 3b).

247

248 *Data analysis*

249 When the Shapiro-Wilk's normality test revealed that the data of both groups (SAL and MUS)
250 on each day (Cond., Test 1, and Test 2) were normally distributed (Figure 5a, 5b, 6a, 6b, 8a, 8b, and 8c),
251 the data were analyzed using a two-way repeated-measures analysis of variance (ANOVA) (Group-Day),
252 with Tukey post-hoc test where appropriate. When the Shapiro-Wilk test failed to show a normal
253 distribution of the data, a nonparametric Mann-Whitney U test was used for the comparison between the
254 groups (Figure 6c, 7a, 7b, 7c, 8c, and 8d). The within-subject comparisons were analyzed by a Friedman
255 test, with a Wilcoxon rank-signed post-hoc test. These statistical analyses were performed using the
256 software OriginPro2016 (Origin Lab, Northhampton, MA, USA).

257

258 *Histological analysis*

259 We used fluorescent-conjugated muscimol (Muscimol, BODIPY™ TMR-X Conjugate, M23400, Thermo
260 Fisher Scientific Inc., Waltham, MA, USA) to visualize the location of the tips of the injector cannulae.
261 After completion of the behavioral experiment, the rats received an overdose of sodium pentobarbital
262 (100 mg/kg). Thirty minutes before the overdosing, fluorescent-conjugated muscimol (250 ng/μl, 0.5
263 μl/side) was microinjected into the BLA. Transcardial perfusion of 0.02 M phosphate-buffered saline was
264 performed followed by fixation with 4% paraformaldehyde (PFA) in 0.1 M phosphate buffer. The brains
265 of the rats were removed and post-fixed in 4% PFA overnight. The brains soaked in 30% sucrose in 0.1 M
266 PB for cryoprotection were cut into 50-μm-thick slices and mounted on gelatin-coated glass slides covers
267 slipped with VECTASHIELD (Vector Laboratories, Burlingame, CA, USA). The locations of the injector
268 tips were evaluated under a fluorescent microscope (AX70-80FLBD, Olympus Corporation, Tokyo,
269 Japan) by an experimentally blind scorer.

270

271

272 **Results**

273 *Location of cannulae tips*

274 Fluorescent images of the brain slices showed tissue labeled with red color around the track of the injector
275 cannulae. We selected the slices having the brightest and largest labeled tissue as the center of the area for
276 injections. Based on the selected slices, we constructed a stereotaxic mapping of the location (Fig. 4)
277 modified from the brain atlas (Paxinos and Watson, 2007). They showed that 8 MUS group rats and 7
278 SAL group rats had the tips of the injector cannulae within the BLA. Eight rats (4 muscimol-injected rats
279 and 4 saline-injected rats) showed that either or both of the injector cannulae were out of the BLA. The
280 behavioral experiment analyses excluded the data of these rats with the injector tips outside the BLA.

281

282 *Behavioral analysis*

283 *Entry*

284 We analyzed the latency of the first entry and the frequency and duration of all entries irrespective of
285 either Entry-Lick or Entry-Stop, to understand how the animals started to approach the bay window.
286 The latency of the first entry showed that the rats reached the bay window about 5 s after the opening of
287 the guillotine door on Conditioning (Table 1). The pairings of the CS with the US did not change the
288 beginning of the first approach on Test 1. In Test 2, while the SAL group showed about two times longer
289 latency than Test 1, the MUS group approached more quickly, though there was no significant group
290 difference.

291 The mean duration of entry—how long the rats stayed at the bay window through the
292 sessions—significantly decreased after the CS-US pairings. The mean duration in the SAL group in Test 2

293 was similar to Test 1 and shorter than the mean duration in the Conditioning stage, while the mean
294 duration in the MUS group for Test 2 was longer than that for Test 1 and was similar for the Conditioning
295 stage. A two-way repeated-measures ANOVA revealed significant main effects for Group ($F_{(1,13)} = 130.71$,
296 $p < 0.001$), Day ($F_{(2,26)} = 10.84$, $p < 0.001$), and Group-Day interaction ($F_{(2,26)} = 7.78$, $p < 0.01$). A Tukey
297 post-hoc test showed a significant group difference in Test 2 ($p < 0.001$). We found within-subject
298 differences in the MUS group (Conditioning vs Test 1, $p < 0.05$; Test 1 vs Test 2, $p < 0.01$).

299 The frequency and total duration of entry decreased after the CS-US pairings in both groups
300 (Table 1), which means the rats decreased approaches to the bay window. The analyses of the total
301 duration of entry using a two-way repeated-measures ANOVA showed a significant main effect for Day
302 ($F_{(2,26)} = 50.20$, $p < 0.001$) but not a main effect of Group ($F_{(1,13)} = 1.89$, $p > 0.05$) and Group-Day
303 interaction ($F_{(2,26)} = 2.08$, $p > 0.05$). Conversely, a Mann-Whitney U test for the frequency of entry
304 revealed a significant group difference for Test 2 ($Z = 2.49$, $p < 0.01$). A Friedman test revealed that the
305 differences in latency among trials were significant in the MUS group ($\chi^2_{(2)} = 9.75$, $p < 0.01$). A Wilcoxon
306 rank-signed post-hoc test indicated the frequency of entry for Test 2 was significantly smaller than that for
307 Conditioning in the MUS group ($p < 0.01$). This suggests that the intra-BLA muscimol injections altered
308 animal approach behavior.

309 We categorized entries into Entry-Lick and Entry-Stop and calculated the proportion of Entry-Lick and
310 Entry-Stop frequencies (Fig. 5b). After the CS-US pairings, the proportion of Entry-Stop increased, while
311 that of Entry-Lick significantly decreased. The MUS group showed a smaller proportion of Entry-Stop
312 and a larger proportion of Entry-Lick than the SAL group for Test 2. We analyzed the data of Entry-Lick
313 and Entry-Stop using a two-way repeated-measures ANOVA, which revealed the main effects for Group

314 ($F_{(1,13)} = 4.82, p < 0.05$) and Day ($F_{(2,26)} = 53.87, p < 0.001$) in both types of entry. The Group-Day
315 interaction was not significant ($F_{(2,26)} = 1.59, p > 0.05$).

316

317 Entry-Lick

318 The CS-US pairings significantly decreased the frequency of Entry-Lick (Fig. 6a). The
319 decreased frequency of Entry-Lick was also observed for Test 2. A two-way repeated-measures ANOVA
320 revealed a significant main effect for Day ($F_{(2,26)} = 50.75, p < 0.001$). We found no significant main effect
321 for Group ($F_{(1,13)} = 0.016, p > 0.05$) and Group-Day interaction ($F_{(2,26)} = 0.153, p > 0.05$). A Tukey
322 post-hoc test showed that the frequency of Entry-Lick for Test 1 and Test 2 was significantly smaller than
323 that for the Conditioning stage in both groups ($p < 0.001$).

324 The mean duration of Entry-Lick was slightly decreased after the pairings of the CS-US (Fig. 6b). The
325 MUS group showed an elevation of the mean duration on Test 2. A two-way repeated-measures ANOVA
326 revealed significant main effects for Group ($F_{(1,13)} = 4.78, p < 0.05$), Day ($F_{(2,26)} = 6.42, p < 0.01$), and
327 Group-Day interaction ($F_{(2,26)} = 6.52, p < 0.01$). A Tukey post-hoc test showed a significant group
328 difference on Test 2 ($p < 0.01$). The mean duration for Test 2 was significantly longer than that for Test 1
329 in the MUS group ($p < 0.01$).

330 We further analyzed the time needed to lick the spout after passing Line 1 as the latency from Line 1 to
331 lick (Fig. 6c)—an index of the speed of head movements in the bay window. The latencies for Test 1 and
332 Test 2 were larger than that for the Conditioning stage. A Mann-Whitney U test showed no significant
333 group difference in every trial (Conditioning, Test 1, and Test 2). A Friedman test revealed that the
334 differences in the latency among trials were significant in both groups (SAL, $\chi^2_{(2)} = 10.57, p < 0.01$; MUS,

335 $\chi^2_{(2)} = 14.25, p < 0.001$). A Wilcoxon rank-signed post-hoc test indicated significant differences between
336 Conditioning and Test 1 (SAL, $p < 0.05$; MUS, $p < 0.05$) and Conditioning and Test 2 (SAL, $p = 0.022$;
337 MUS, $p < 0.05$). We also found a significant difference between Test 1 and Test 2 in the MUS group ($p <$
338 0.001).

339

340 Entry-Stop

341 The CS-US pairings increased the frequency of Entry-Stop (Fig. 7a). For the SAL group, the frequency in
342 Test 2 was larger than in Test 1, while the MUS group showed a decrease in the frequency. We analyzed
343 data of the total frequency (sum of frequency at Line 1, 2, and 3) and the frequency at each line using a
344 Mann-Whitney U test, which revealed significant group differences in total frequency for Test 2 ($Z = 3.01$,
345 $p < 0.001$). It also showed group differences in the frequency at Line 1 for Test 1 ($Z = 2.04, p < 0.05$) and
346 Test 2 ($Z = 2.05, p < 0.05$). The within-subject comparisons using a Friedman test showed significant
347 differences among trials in the SAL group (Line 3, $\chi^2_{(2)} = 7.36, p < 0.05$) and the MUS group (total, $\chi^2_{(2)}$
348 $= 6.94, p < 0.05$). A Wilcoxon rank-signed post-hoc test demonstrated significant differences in the
349 frequency at Line 3 between Conditioning and Test 2 in the SAL group ($p < 0.05$) and the total frequency
350 between Conditioning and Test 1 in the MUS group ($p < 0.05$).

351 As the MUS group showed a lower frequency of Entry-Stop than the SAL group in Test 2, we divided the
352 session into a first and second half (Fig. 7b). MUS group demonstrated a significantly smaller frequency
353 of Entry-Stop than SAL group in the first half ($Z = 2.84, p < 0.01$). This suggests that the intra-BLA
354 muscimol injections significantly inhibited Entry-Stop on the expression of CTA.

355 The mean duration of Entry-Stop at Line 2 and 3 on Test 1 increased after the CS-US pairings (Fig. 7c).
356 Both groups showed a higher mean duration for Entry-Stop at Line 3 than for Conditioning. We analyzed
357 the differences in mean duration at each line between the SAL and MUS groups using a Mann-Whitney U
358 test. We found no significant group difference, indicating that the intra-BLA muscimol injections had no
359 effect on staying time at the bay window. A Friedman test revealed significant differences in the mean
360 duration among trials in both groups (SAL, $\chi^2_{(2)} = 8.86, p < 0.05$; MUS, $\chi^2_{(2)} = 2.28, p < 0.05$). A
361 Wilcoxon rank-signed post-hoc test showed a significant difference between Conditioning and Test 1 in
362 the MUS group ($p < 0.05$) and between Conditioning and Test 2 in both groups ($p < 0.05$, respectively).
363
364 Licking behavior

365 The total number of licks during the session remarkably declined after the CS-US pairings (Fig. 8a). In
366 Test 2, the MUS group showed more licks than the SAL group. A two-way repeated-measures ANOVA
367 revealed significant main effects for Day ($F_{(2,26)} = 149.43, p < 0.001$) and Group-Day interaction ($F_{(2,26)} =$
368 $9.28, p < 0.001$). The main effect for Group was not significant ($F_{(1,13)} = 1.60, p > 0.05$). A Tukey post-hoc
369 test showed a significant group difference for Test 2 ($p < 0.01$). We also found the total number of licks in
370 Test 1 was significantly lower than in that in Conditioning for both the groups ($p < 0.001$). In Test 2, the
371 SAL group showed a slight increase in the total number of licks (vs Conditioning, $p < 0.001$; vs Test 1,
372 n.s.), while the MUS group had less lick number than in Conditioning ($p < 0.001$) but larger than Test 1 (p
373 < 0.001). These results indicate that the intra-BLA muscimol injections increased CS licking but did not
374 elevate it to the level before the establishment of CTA (Conditioning).

375 After the CS-US pairings, the burst number largely decreased (Fig. 8b). There was no clear group
376 difference in Test 2. A two-way repeated-measures ANOVA revealed significant main effects for Day
377 ($F_{(2,26)} = 35.71, p < 0.001$). The main effect for Group and Group-Day interaction were not significant
378 ($F_{(1,13)} = 0.73, p = 0.408$; $F_{(2,26)} = 1.99, p > 0.05$). A within-subject comparison among the trials using a
379 Tukey post-hoc test showed significant differences in the number of bursts between Conditioning and Test
380 1 ($p < 0.001$) and Conditioning and Test 2 ($p < 0.001$).

381 The burst size also decreased after the CS-US pairings (Fig. 8c). In Test 2, the MUS group showed a
382 higher burst size. A Mann-Whitney U test revealed a significant group difference for Test 2 ($Z = -2.60, p$
383 < 0.01). A Friedman test showed significant differences among trials in the SAL group ($\chi^2_{(2)} = 10.57, p <$
384 0.01) and the MUS group ($\chi^2_{(2)} = 13.00, p < 0.01$). A Wilcoxon rank-signed post-hoc test demonstrated a
385 significantly small burst size for Tests 1 and Test 2 compared to Conditioning in the SAL group ($p < 0.05$).

386 On the other hand, the MUS group displayed a significant decrease in burst size for Test 1 ($p < 0.01$) and
387 increase for Test 2 ($p < 0.01$).

388 For a more detailed analysis of burst number and size, we plotted the size of all bursts in all the rats for
389 Test 2 (Fig. 9a, b). The MUS group displayed a significantly larger number of bursts with 100 licks more
390 than the SAL group (Fig. 9c; $p < 0.05$). These results suggest that the intra-BLA muscimol injections
391 increased burst licking to the CS.

392 The CS-US pairings altered the proportions of the inter-lick interval (Fig. 8d). The rats exclusively
393 showed a < 200 ms ILI in Conditioning. In Test 1, ILI decreased, whereas IBI ($200 \leq \text{ILI} < 500$ ms), ICI
394 ($500 \leq \text{ILI} < 1000$ ms), and pause ($1000 \text{ ms} \leq \text{ILI}$) increased. The SAL group showed a similar proportion
395 of intervals with Test 1. Conversely, the MUS group had similar proportions with Conditioning and

396 demonstrated smaller proportions of ILI, IBI, and Pause than in Test 1. We analyzed group differences in
397 the proportion of each type of interval using a Mann-Whitney U test. The MUS group showed
398 significantly larger ILI ($p < 0.05$) and significantly smaller ICI and pause ($p < 0.01$, both) than the SAL
399 group in Test 2. A Friedman test showed significant trial differences in ILI (SAL, $\chi^2_{(2)} = 10.57, p < 0.01$;
400 MUS, $\chi^2_{(2)} = 13.00, p < 0.01$), ICI (SAL, $\chi^2_{(2)} = 11.14, p < 0.01$; MUS, $\chi^2_{(2)} = 7.75, p < 0.05$), and Pause
401 (SAL, $\chi^2_{(2)} = 10.57, p < 0.01$; MUS, $\chi^2_{(2)} = 12.00, p < 0.01$). A Wilcoxon rank-signed post-hoc test
402 demonstrated significant differences in ILI, ICI, and pause between Conditioning and Test 1 ($p < 0.05$,
403 all), Conditioning and Test 2 in the SAL group ($p < 0.05$, all). On the other hand, the MUS group showed
404 no differences between Conditioning and Test 2, and significant differences in ILI and Pause between Test
405 1 and Test 2 ($p < 0.01$, respectively). These results indicate that the establishment of CTA induced a
406 longer inter-lick interval, and that intra-BLA muscimol injections made the inter-lick interval shorter.

407

408 **Discussion**

409 We examined the effects of muscimol microinjections into the BLA on approach and licking behaviors in
410 rats during CTA retrieval. We found that intra-BLA muscimol microinjection changes animal behavior,
411 leading to an increase in the mean duration but not the frequency of Entry-Lick, a decrease in the
412 frequency but not the mean duration of Entry-Stop, an augmentation of lick number and burst size, and a
413 reduction in the inter-lick interval.

414

415 *Effects of muscimol injections on Entry-Lick*

416 Intra-BLA muscimol increased the mean duration of Entry-Lick (Fig. 6b) but did not affect Entry-Lick
417 frequency. The muscimol injections also induced a larger total lick number during the 30 min test session
418 (Fig. 8a). As shown in Fig. 2, the proportion of licking behavior for each entry (size of blue raster) was
419 larger in the MUS group than in the SAL group. This indicates that muscimol injections induce relatively
420 long-lasting licking behavior resulting in longer duration of Entry-Lick.

421 BLA inactivation reduces approach behavior to positive rewards, such as food and drugs (Fuchs and See,
422 2002; Kantak et al., 2002; McLaughlin and See, 2003; Fuchs et al., 2005; Rizos et al., 2005; McLaughlin
423 and Floresco, 2007; Savage et al., 2007; Peters et al., 2008; Gabriele and See, 2010; Jones et al., 2010;
424 Chaudhri et al., 2013; Parkes and Balleine, 2013; Parkes et al., 2016). Conversely, the entrance of the rat's
425 head into the bay window in this study was followed by licking to the aversive CS. Since the approach
426 behavior was linked to a negative reward (e.g. CS), intra-BLA muscimol had no effect on the frequency
427 of Entry-Lick. This also suggests that BLA inactivation does not cause changes in drinking motivation
428 and locomotor activities.

429

430 *Effects of muscimol on Entry-Stop*

431 Since intra-BLA muscimol decreased the frequency of Entry-Stop for Test 2 (Fig. 7a, b), we examined
432 why CTA establishment increases the mean duration of Entry-Stop, as shown in Fig. 7c.

433 Several reports used a CTA paradigm as a model for approach-avoidance conflict (Weinberg et
434 al., 1982; Brett et al., 1986; Ervin and Cooper, 1988). A conflict has been hypothesized to occur when two
435 or more incompatible response tendencies compete (Miller, 1944). Simultaneous tendencies to approach
436 and avoid the goal produce a conflict situation. In a single-bottle CTA study, when an animal is

437 water-deprived and provided only with a conditioned aversive taste solution, the animal is in a presumed
438 conflict situation between thirst (approach tendency: motivation to drink) and disgust (avoidance
439 tendency: inhibition to drink) (Ervin and Cooper, 1988). Since we put a spout in front of the bay window
440 and the rats were water-deprived, the rats possibly were in an approach-avoidance conflict situation in
441 Test 1 and Test 2.

442 Miller (1944) described that the degree of conflict depends on the relative strengths of the competing
443 tendencies. Thus, maximum conflict arises when these strengths are equal. The animal's proximity to the
444 goal and drive state affects the strengths. The frequency of stops at Line 3 (Fig. 7a) and mean duration of
445 stops at Line 2 and 3 in Test 1 (Fig. 7c) were larger than Conditioning, indicating that the rats reached
446 close to the spout (to Line 3) but did not lick the spout even when they were motivated to drink fluid due
447 to water deprivation after the CS-US pairings. Therefore, we assume that the Entry-Stop in Test 1 and Test
448 2 reflects a maximum approach-avoidance conflict situation.

449 The proportion of frequency for Entry-Stop in entire entry events for the MUS group was lower than that
450 for the SAL group in Test 2 (Fig. 5b). The SAL group tended to show a larger frequency for Entry-Stop in
451 Test 2 than in Test 1, while the MUS group demonstrated no difference between Test 1 and Test 2. The
452 frequency of Entry-Stop in the first half of the session in the MUS group was significantly smaller than
453 the SAL group (Fig. 7b). These results suggest that intra-BLA muscimol reduces avoidance tendency.

454 Since there were no changes in the frequency of Entry-Lick in the MUS group (Fig. 6a), the intra-BLA
455 muscimol injections did not alter the approach tendency. Therefore, the reduction in avoidance tendency
456 is considered to attenuate approach-avoidance conflict on CTA retrieval.

457 Intra-BLA muscimol had no effect on the mean duration of Entry-Stop (Fig. 7c) unlike on the frequency,
458 indicating that the MUS group, similar to the SAL group, stopped and stayed in the bay window for a
459 short duration. Since the lidocaine-reversible inactivation of the BLA was found not to affect spontaneous
460 or basal locomotor activity (Woods and Ettenberg, 2004), the significant reduction in the frequency of
461 Entry-Stop in the MUS group was unlikely to be associated with to the alteration in locomotor activities.
462 The administration of the anti-emetic ondansetron attenuates aversive taste reactions to the intra-orally
463 infused CS, which is an index of decreased palatability, but it does not affect the consumption of the CS
464 (Limebeer and Parker, 2000). They argue that animal behaviors on CTA retrieval comprise decreased
465 palatability and avoidance. Therefore, the reduction in avoidance tendency in the MUS group in this study
466 was possibly due to avoidance attenuation to the CS.

467

468 *Effects of muscimol on licking behavior*

469 Intra-BLA muscimol increased the total number of licks (Fig. 8a), indicating increased CS consumption.
470 Midazolam microinjections into the BLA increased the intake of the conditioned aversive sucrose
471 (Yasoshima and Yamamoto, 2005). Midazolam does not directly activate GABA_A receptor but enhances
472 the effect of GABA on GABA_A receptors, facilitating the opening of the Cl⁻channel. Thus, midazolam is
473 considered to have a neural inhibitory effect. The administration of an AMPA receptor blocker NBQX
474 into the BLA also increases the CS intake (Garcia-Delatorre et al., 2014). These findings indicate that the
475 BLA inactivation might impair the suppression of the consumption of a CS on CTA retrieval.
476 The establishment of CTA decreases the burst number and size (Baird et al., 2005; Rebecca Glatt et al.,
477 2015). The significantly reduced burst number (Fig. 8b) and burst size (Fig. 8c) in Test 1 demonstrates

478 that the rats in this study acquired a robust aversion. Although the group difference in the burst number in
479 Test 2 was not significant, the burst size in the MUS group was significantly larger than that in the SAL
480 group. These results suggest that intra-BLA muscimol attenuated aversion to the CS.

481 The plots of burst size in Test 2 (Fig. 9a, b) demonstrated a higher number of bursts with over 100 licks in
482 the MUS group. These results suggest that intra-BLA muscimol tends to increase burst size. Higher
483 concentrations of sucrose produce larger burst size (Davis and Smith, 1992; Davis and Perez, 1993;
484 Spector et al., 1998), while conditioned aversive taste reduces burst size (Dwyer et al., 2008; Swick et al.,
485 2015). Naturally, aversive quinine also produces smaller burst size than water (Spector and St John, 1998).
486 Therefore, the burst size in the MUS group also suggests the attenuated aversion to CS by BLA
487 inactivation.

488 The proportion of ILI (< 200 ms) decreased, and that of IBI (200-500 ms), ICI (500-100 ms), and Pause
489 (> 1000 ms) increased after the CS-US pairings (Fig. 8d). A previous study showed an increased
490 proportion of longer inter-lick interval (Baird et al., 2005), supporting the argument that the rats in Test 1
491 showed aversion to the CS. On Test 2, the SAL group demonstrated proportions of IBI, ICI, and Pause
492 similar to those for Test 1, whereas the proportions in the MUS group was similar to that in the
493 Conditioning stage. This indicates that attenuation of aversion to the CS by intra-BLA muscimol
494 injections was also identified through the reduction in the inter-lick interval.

495

496 *The Efferent projections from the BLA on CTA retrieval*

497 CTA retrieval activates neuronal projections from the BLA to the NAcC (Inui et al., 2013) and NAcC
498 neurons (Yasoshima et al., 2006) and changes the neurotransmission in the NAcC (Mark et al., 1991;

499 Mark et al., 1995). CTA retrieval also activates GABAergic transmission from the NAcC to the VP (Inui
500 et al., 2007; Inui et al., 2009). These findings suggest that the BLA-NAcC-VP neural circuits mediate the
501 decreased palatability on CTA retrieval. Further, we have previously shown that CTA retrieval activates
502 the neuronal projections from the BLA to BNST (Inui et al., 2013), but the role of the BNST in CTA
503 retrieval is still unclear. Since the BNST is involved in fear responses (Davis, 1992; Duvarci et al., 2009;
504 Walker et al., 2009), the BLA-BNST pathway may be involved in the avoidance tendency on CTA
505 retrieval.

506

507

508 **Conclusion**

509 In this study, we developed an experimental chamber for analyzing the temporal sequence of the
510 behaviors of rat approaches to a spout and CS licking behaviors. The results showed that microinjections
511 of muscimol into the BLA reduced avoidance to the CS. Muscimol injections also increased total CS
512 licking, and lick microstructural analysis demonstrated that the attenuation of aversion increased CS
513 licking. These results reveal that the BLA essentially contributes to CTA retrieval by simultaneous
514 processing of avoidance and palatability to the CS.

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669

670 **Figure legend**

671 *Figure 1: The experimental chamber*

672 A. The box was separated into a start box and an arena. Animals were put in the start box at the start of
673 the trials. The animals were allowed to enter the arena through the door opened and closed by an
674 experimenter. There was a bay window on the opposite side of the door in the arena. B. The photograph
675 shows a rat approach the bay window after the opening of the door at the start of a trial. C-E. A diagram
676 and photograph of the bay window. The rats put their heads into the bay window as shown in (D) to lick a
677 spout in front of the bay window. Infrared LEDs below and sensors above the bay window detected the
678 entry of the heads. The metal spout was connected with a touch sensor via a metal clip and wire.

679

680 *Figure 2: Raster plots of the responses of the passing and touch sensors in each animal on Test 2*

681 The red, yellow, and green vertical lines indicate Line 1 (far from the spout), Line 2 (middle of the bay
682 window), and Line 3 (close to the spout), respectively. The red color bars indicate that the rat's head (tip
683 of nose) stopped at the entrance of the bay window. Both the red and yellow color bars indicate the stop at
684 the middle of the bay window, and the sets of the red, yellow, and green bars indicate the stop just before
685 the spout. The plots with a gray background in the saline group are enlarged in Figure 3 for explaining
686 behavioral analyses.

687

688 *Figure 3: Scheme for behavioral analyses*

689 A. The analysis of the entrance of the rat's heads into the bay window (Entry). Top: The blocks containing
690 all the color bars indicate Entry-Lick, while the ones containing all the colors without blue indicate

691 Entry-Stop. Bottom: Illustrations showing the animal's behavior on Entry-Lick and Entry-Stop. B.
692 Schematic diagram of the licking pattern. The vertical lines indicate licks. ILI, inter-lick interval; IBI,
693 inter-burst interval; ICI, inter-cluster interval.

694

695 *Figure 4: Histological findings.*

696 The locations of the injection cannulae tips were reconstructed based on the Paxinos and Watson brain
697 atlas (2007). Orange circles, muscimol-injected rats; black squares; saline-injected rats.

698

699 *Figure 5: The intra-BLA muscimol altered Entry*

700 A. The mean duration of all Entry in the saline-injected (SAL) and muscimol-injected (MUS) groups on
701 Conditioning, Test 1, and Test 2. After the CS-US pairings, the mean duration of Entry decreased in both
702 groups. The microinjections of muscimol into the BLA on Test 2 impaired the decline in the mean
703 duration of Entry. B. The proportion of the frequency of Entry-Lick and Entry-Stop. The conditioning
704 decreased Entry-Lick and increased Entry-Stop. The MUS group showed smaller proportion of
705 Entry-Stop and larger Entry-Lick than the SAL group on Test 2. $***p < 0.001$ (vs. SAL.), $*p < 0.05$, $^\dagger p <$
706 0.05 (vs. Conditioning), $^{ss}p < 0.01$ (vs. Test 1).

707

708 *Figure 6: The intra-BLA muscimol decreased the mean duration of Entry-Lick*

709 A. The frequency of Entry-Lick in the saline-injected (SAL) and muscimol-injected (MUS) groups on
710 Conditioning, Test 1, and Test 2. The CS-US pairings considerably decreased the frequency of Entry-Lick.
711 Both the SAL and MUS groups showed suppressed frequency of Entry-Lick on Test 2. There was no

712 group difference. B. The mean duration of Entry-Lick. The rats showed a slightly shorter mean duration
713 of Entry-Lick on Test 1 than Conditioning. On Test 2, the SAL group showed the same level of mean
714 duration with Test 1, while the MUS group demonstrated significantly longer mean duration than Test 1
715 and the SAL group on Test 2. C. The latency from the cut of Line 1 to lick. The CS-US pairings
716 significantly increased latency. The microinjections of muscimol into the BLA did not alter the latency in
717 Test 2. $**p < 0.01$ (vs. SAL), $†††p < 0.001$, $†p < 0.05$ (vs. Conditioning), $^{ss}p < 0.01$, $^sp < 0.05$ (vs. Test 1).

718

719 *Figure 7: The intra-BLA muscimol decreased the frequency of Entry-Stop*

720 A. The frequency of Entry-Lick in the saline-injected (SAL) or muscimol-injected (MUS) group in
721 Conditioning, Test 1, and Test 2. The CS-US pairings increased Entry-Stop. On Test 2, the MUS group
722 showed significantly smaller stops than the SAL group. B. The frequency of Entry-Stop during the first
723 and last half of Test 2. The muscimol-injected group showed smaller frequency of the stop on both the
724 time windows. There was a significant group difference for the first half. C. Mean duration of Entry-Stop
725 at each line. The CS-US pairings significantly increased mean duration at Line 2 and 3. The mean
726 duration at Line 2 decreased in both groups for Test 2 and increased at Line 3 for Test 2. However, there
727 were no differences between the groups. C, Conditioning; T1, Test 1; T2, Test 2. $**p < 0.01$, $*p < 0.05$ (vs.
728 SAL), $†p < 0.01$ (vs. Conditioning).

729

730 *Figure 8: The intra-BLA muscimol attenuated aversion to the CS*

731 A. The total number of licks in the saline-injected (SAL) and muscimol-injected (MUS) groups on
732 Conditioning, Test 1, and Test 2. The CS-US pairings significantly decreased the total number of licks.

733 The MUS group showed larger CS licking than the SAL group on Test 2. B. The number of bursts. The
734 CS-US pairings decreased the burst number. There was no group difference for Test 2. C. The size of the
735 burst. The CS-US pairings decreased the burst size. The MUS group demonstrated a larger burst size than
736 the SAL group for Test 2. D. The proportion of inter-lick interval. The CS-US pairings increased the
737 proportions of IBI, ICI, and Pause, which were smaller in the MUS group than in the SAL group. $**p <$
738 0.01 , $*p < 0.05$ (vs. SAL), $^{+++}p < 0.001$, $^{++}p < 0.01$, $^{\dagger}p < 0.05$ (vs. Conditioning), $^{sss}p < 0.001$, $^{ss}p < 0.01$,
739 $^s p < 0.05$ (vs. Test 1).

740

741 *Figure 9: The intra-BLA muscimol increased larger burst size*

742 Plots for all burst sizes in the saline-injected (SAL) and muscimol-injected (MUS) groups for Test 2. A.

743 The size of almost all the bursts was less than 100 licks in the SAL group. B. The MUS group showed a

744 larger number of lick bursts containing over 100 licks. C. There was a significant group difference in the

745 average number of >100 lick burst size. $*p < 0.05$ (vs. SAL).

746

747 **Tables**

748 *Table 1*

749 Latency, frequency, and total duration of Entry

750 $**p < 0.01$, $*p < 0.05$ (vs. SAL), $^{\dagger}p < 0.01$ (vs. Conditioning).

Figure 1

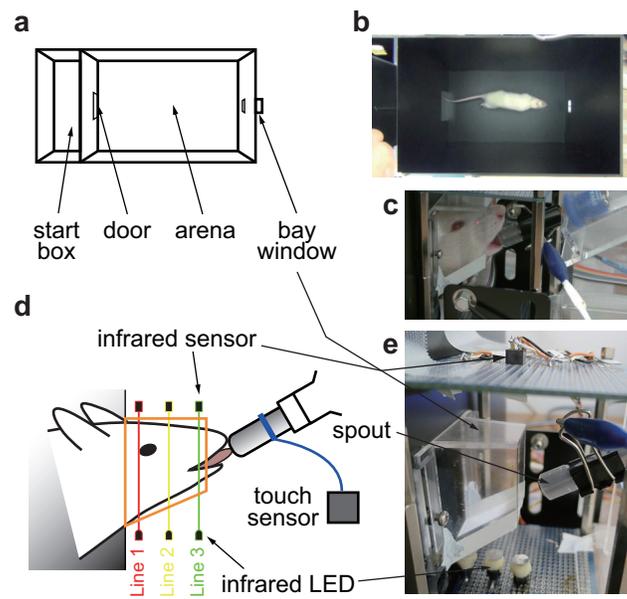


Figure 2

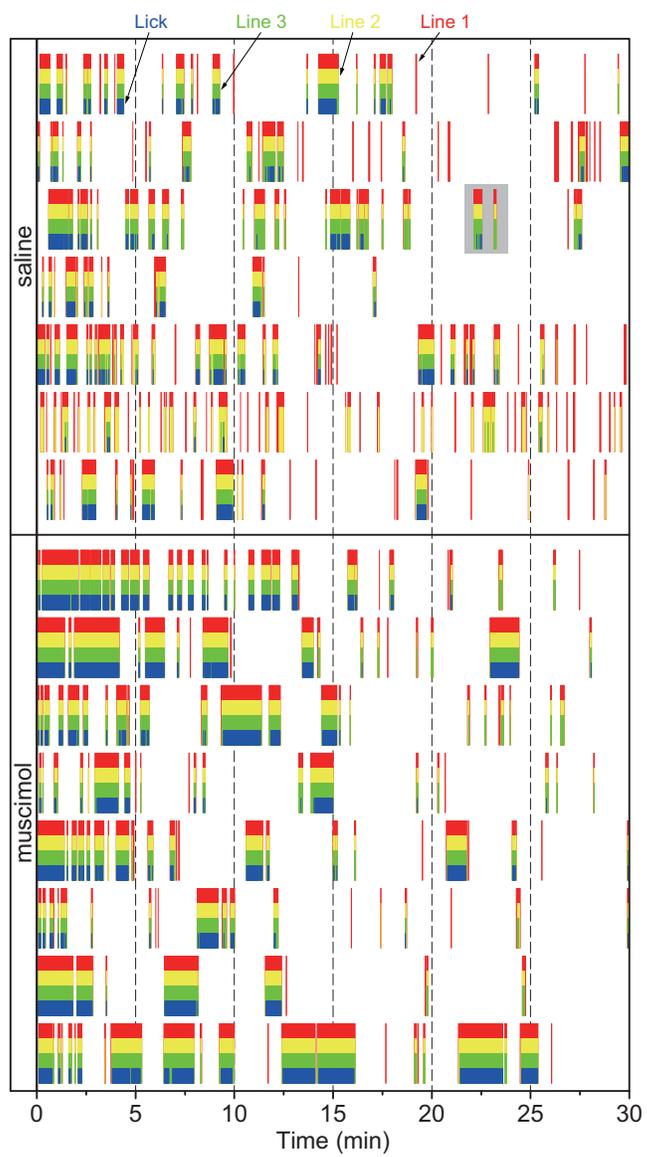


Figure 3

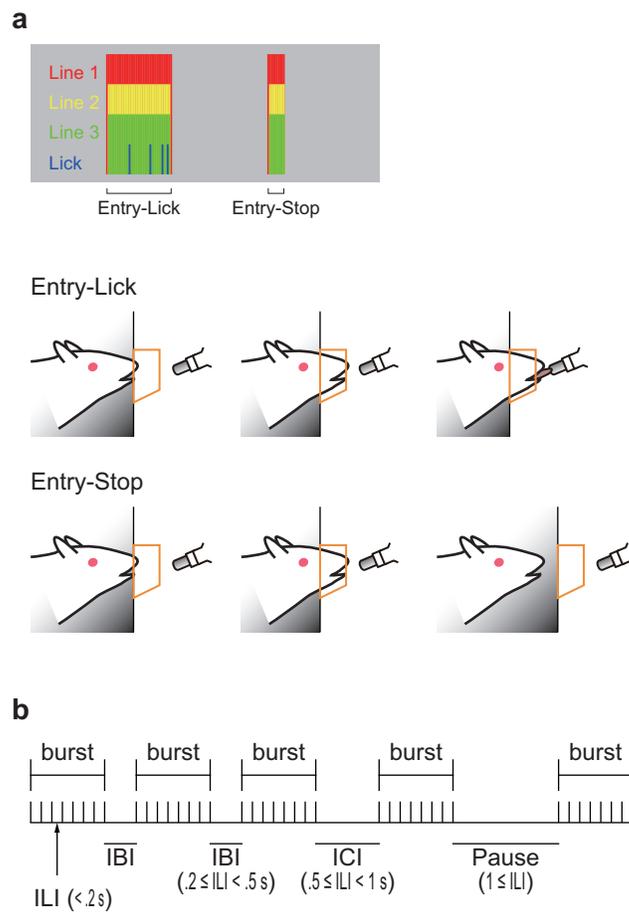


Figure 4

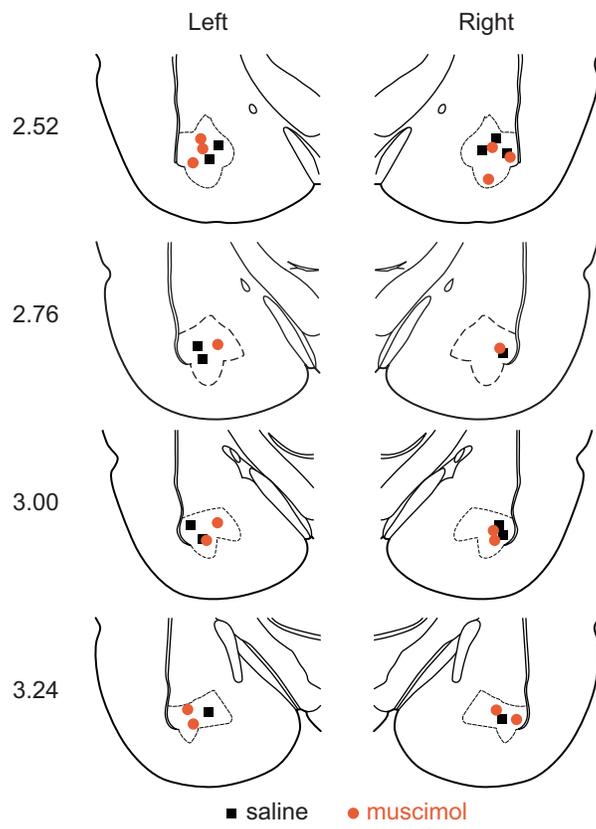


Figure 5

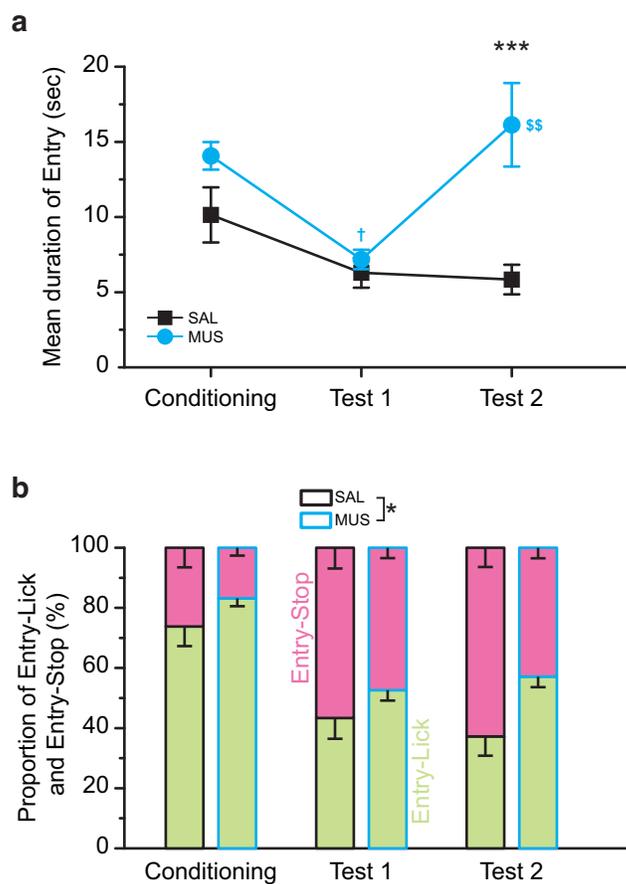


Figure 6

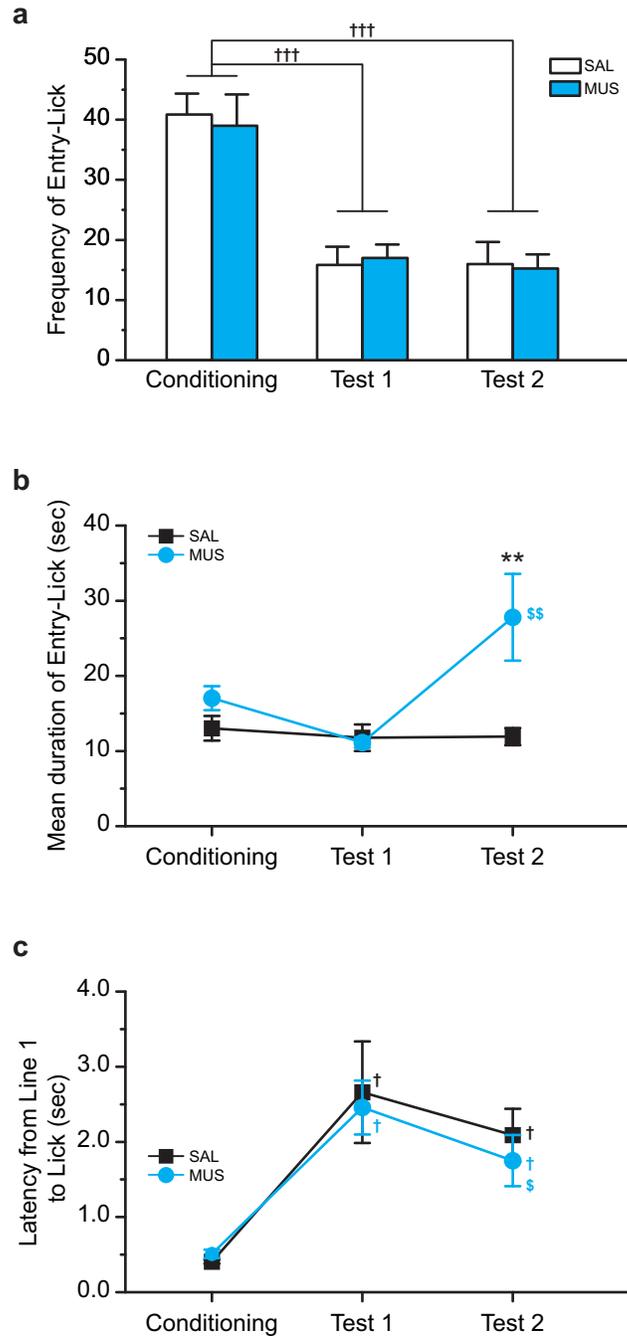


Figure 7

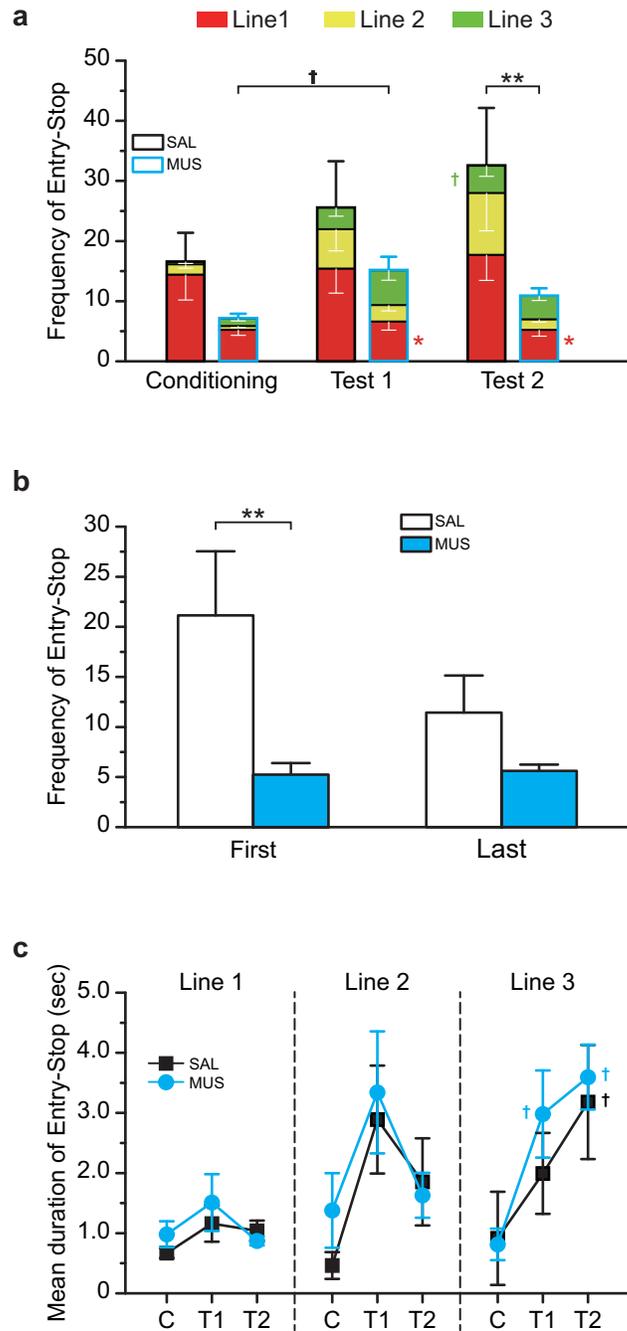


Figure 8

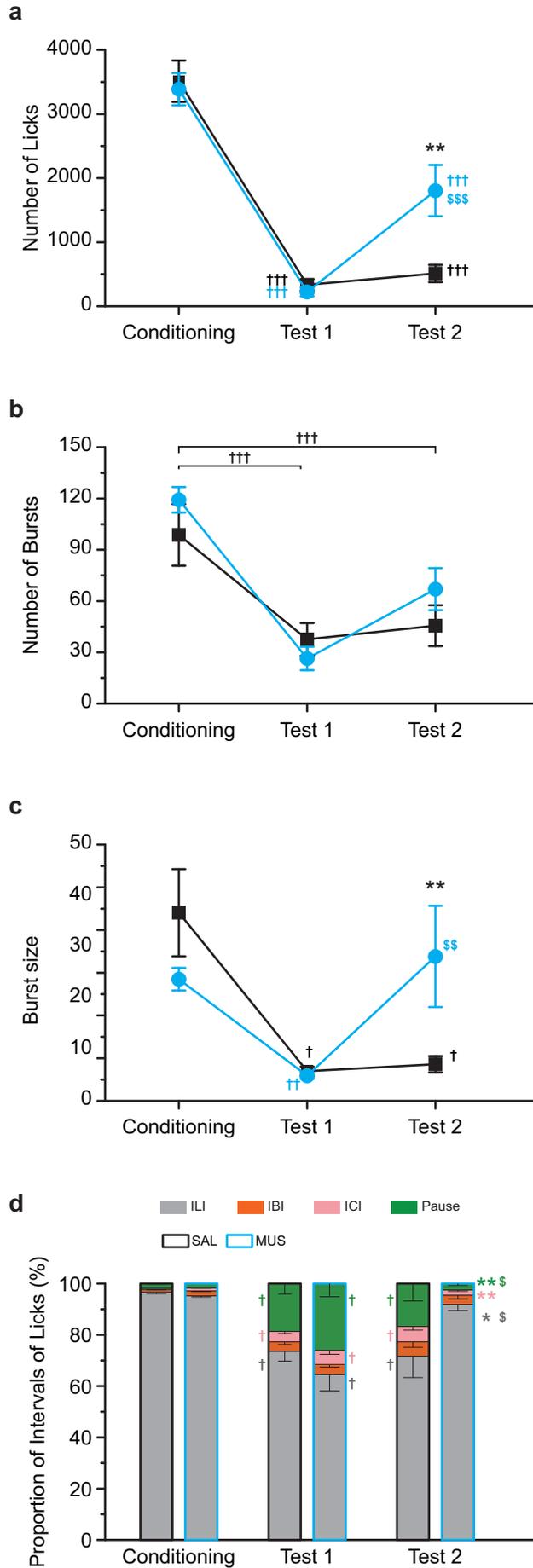


Figure 9

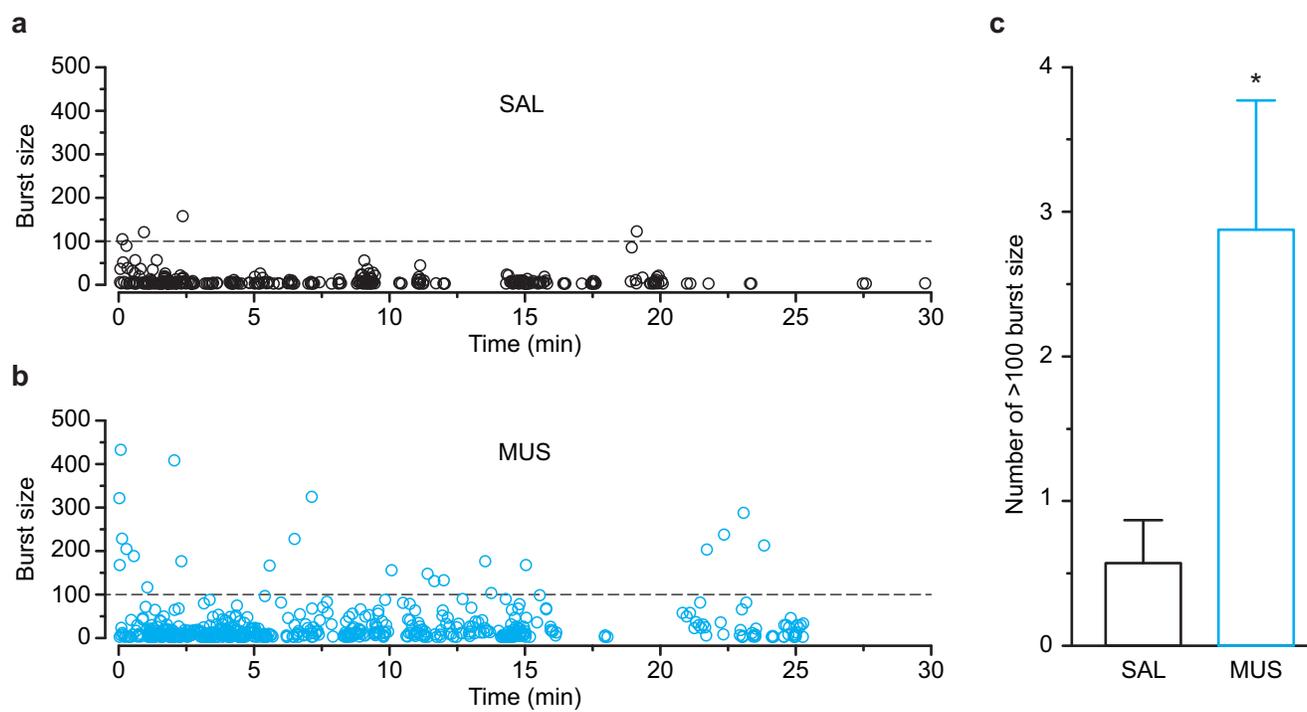


Table 1

	Conditioning		Test 1		Test 2	
	SAL	MUS	SAL	MUS	SAL	MUS
Latency of 1st Entry	4.66 ± 0.84	5.91 ± 0.63	8.73 ± 0.93	4.18 ± 0.94	13.34 ± 4.27	*4.46 ± 0.87
Frequency of Entry	57.43 ± 5.89	46.13 ± 4.86	41.43 ± 7.30	32.13 ± 4.02	67.86 ± 21.4	††**26.13 ± 2.91
Total duration of Entry	534.6 ± 64.14	625.93 ± 35.99	243.19 ± 41.56	239.63 ± 41.57	255.67 ± 32.15	393.18 ± 62.49